

International Transaction Journal of Engineering, Management, & Applied Sciences & Technologies

http://TuEngr.com



# **Optimization of Enzymatic Clarification from Corncob**

Sininart Chongkhong <sup>a\*</sup>, and Woraluk Kongjindamunee <sup>a</sup>

<sup>a</sup> Department of Chemical Engineering Faculty of Engineering, Prince of Songkla University, THAILAND

ARTICLEINFO	A B S T RA C T
Article history: Received 16 August 2013 Accepted 06 December 2013 Available online 09 December 2013	A major content that was 22.76% total carbohydrate of the corncob could be simply hydrolyzed into reducing sugars by using alpha-amylase. The clarification process using alpha-amylase was optimized by response surface methodology (RSM) in this work.
<i>Keywords</i> : Alpha-amylase; Glucose content; Hydrolysis; Central composite design;	Independent variables including: enzyme amount of 0.05-0.2 %w, time of 60-240 min and temperatures of 80-100 °C were investigated. Their effects were found on the reducing sugar (Glucose content) by a second order central composite design (CCD). The optimum condition was 0.2 %w alpha-amylase, 87.6 °C for 150 min. It could provide the highest amount of 6.21 g/L glucose content in the clarified product.
	©2014 INT TRANS J ENG MANAG SCI TECH.

# 1. Introduction

Ethanol is an alternative energy source as the clean and safe transportation fuels that can be produced domestically in response today's high-energy demand. This renewable energy has been interesting and rapidly developing to be used for substituting on fossil fuels and reducing pollution. Agricultural residues are used economically as raw materials for the ethanol production (Liu *et al.*, 2010; Chena *et al.*, 2007). The raw materials can be conveniently classified into three types: (i) sugars such as sugar beet, sweet sorghum and sugar cane, (ii) easily degradable carbohydrates such as corn, rice, wheat barley and corncob, (iii) cellulose such as rice bran, rice straw, wood chips, sawdust and waste from industries (Kreuger *et al.*, *a.*)

2011; Zhu *et al.*, 2006). The production steps are pretreatment, hydrolysis and fermentation, respectively. The pretreatment are necessary for increasing the porosity of the materials that are active to next steps. Then the hydrolysis, cellulose and/or carbohydrate molecules are converted into reducing sugars or broken down into corresponding monomers. After that the fermentation is carried out to transform the reducing sugars (fermentable sugars) into ethanol (Balata *et al.*,2008). Corncob is a cheap raw that can give a suitable reducing sugar (glucose) content for ethanol production. The components per 100 g of corncob are 22.76 g total carbohydrate, 7.11 g crude fiber, 1.17 g crude protein, 0.15 g fat, 0.44 g ash and 75.48 g moisture (Agro-Industry department center for export, 2011). The alpha-amylases breaks down the long-chain carbohydrates by acting at random locations of the carbohydrate chain. In this process, the pH is adjusted to be about 6.0 - 6.5 and the reaction is performed for about 2 hours at 95°C (Aiyer, 1995; Das *et al.*, 2011; Reed and Nagodawithana, 1995). The optimum condition for the hydrolysis of prebiotic extracted jackfruit seeds using alpha-amylase enzyme was 0.17 %w enzyme amount at 80 °C for 240 min. The highest reducing sugar content in the product was 3.04 g/L (Bancha *et al.*, 2011).

Response surface methodology (RSM) is a statistical technique to identify the effect of individual variable for the optimization of multivariable system. It is widely used in optimizing the bioprocesses by the statistical experimental design method. This method can be employed to determine the optimum processes i.e. pretreatment, hydrolysis and fermentation. In addition, it can enhance production yield, reduce process variability, save time and cost (Wang *et al.*, 2008; Bandaru *et al.*, 2006).

Crucial factors (alpha-amylase amount, time and temperature) for clarification process from corncob in this work would be optimized by RSM to obtain the highest glucose content in the product.

# 2. Materials and Methods

## 2.1 Materials and chemicals

The corncob of sweet corn, sugar specie, was obtained from a local market in Hat-Yai, Songkhla province, Thailand. The composition of the corncob is shown in Table 1. Alpha-amylase from *Aspergillus oryzae* was purchased from the Sigma-Aldrich company. Dinitrosalicylic solution (DNS) was used for the analysis of the glucose (reducing sugar) in the products (Miller, 1959). DNS was the mixture of 1% dinitrosalicylic acid, 0.2% phenol, 0.05% sodium sulfite, 1% sodium hydroxide and 20% sodium potassium tartrate that were a laboratory

grade.

**Test Items** Test Method Results AOAC Protein 1.17 % (Kjeldahl Method) AOAC Crude Fat 0.15 % (Soxhlet Extraction Method) AOAC Moisture 75.48 % (Loss on Drying at 95-100 °C Method) Ash AOAC 0.44 % AOAC Crude Fiber 7.11 % (Fritted Glass Crucible Method) Total Calculation 22.76 % Carbohydrate 97.07 kcal Calculation Energy Modified Phenol Sulfuric Method Total Sugar 3.71 % Reducing sugar Modified dinitrosalicylic acid method 1.68 g/L

Table 1: The components of the corncob content.

## 2.2 Pretreatment and pre-hydrolysis (Clarification)

The corncob was firstly cut into small pieces and crushed to be about 2 mm particle size. The 20 g crushed corncob and 100 mL clean water were put into 250 ml screw-capped bottles, and added with 0.05-0.2 %w alpha-amylase. An initial pH was adjusted to be 6.0 by ammonia solution. Then the bottles were immersed in an oil bath at a studied temperature in the range of 80-100 °C for a heating time in the range of 60-240 min with a constant shaking rate of 80 rpm. After that, the clarified products were separated by a fabric filter to get the clear liquid phase product before the analysis of reducing sugar content by a UV-vis spectrophotometer.

## 2.3 Analytical method

DNS method using a double beam UV-Vis spectrophotometer (Model HP 8453) with UV-Visible Chem-Station software was used to analyze the reducing sugar that was assayed in term of glucose. The reflective light was measured at 520 nm on the spectrophotometer (Chongkhong et al., 2012).

## 2.4 Experimental design and optimization

Central composite design (CCD) was employed to assign important parameters for investigation. Time ( $X_1$ , min), temperature ( $X_2$ , °C) and alpha-amylase amount ( $X_3$ , %w) were chosen as the independent variables that are shown in Tables 2 and 3. The reducing sugar concentration in the product (Y, g/L) was the dependent output variable. For statistical calculation, the variables were coded according to Equation (1).

$$x_i = \frac{(X_i - \bar{x}_i)}{\Delta x_j}, i = 1, 2, 3, \dots, k$$
 (1),

where  $x_i$  and  $X_i$  are the dimensionless value and the real value of the independent variable,  $\overline{x_i}$  is the real value of the independent variable at the center point and  $\Delta x_i$  is the step change.

Variables	Coded levels					
	-1.68	-1	0	1	1.68	
Time (min)	60	95	150	205	240	
Temperature (°C)	80	85	90	95	100	
Alpha-amylase (%w)	0.05	0.08	0.13	0.17	0.2	

**Table 2:** Independent variables for the experimental design.

<b>Table 3:</b> The central composite design matrix employed for the three independent variables
(Actual values are given in Table 2)

Ъ	(Actual values are given in Table 2)						
Run no.	$X_1$	X <sub>2</sub>	X <sub>3</sub>				
1	1	-1	-1				
2	0	0	0				
3	-1	1	1				
4	-1.68	0	0				
5	0	-1.68	0				
6	0	0	0				
7	-1	-1	-1				
8	0	0	1.68				
9	1.68	0	0				
10	-1	1	-1				
11	-1	-1	1				
12	0	1.68	0				
13	0	0	-1.68				
14	1	-1	1				
15	1	1	-1				
16	1	1	1				
17	0	0	0				

The 17 experiments (N) were estimated by  $N = 2^n+2n+n_0$  that their operating conditions were performed in Table 3. This design consists of the following three portions:

(1) A complete  $2^n$  factorial design, when n is number of test variables.

(2)  $n_0$  center point ( $n_0 \ge 1$ ).

(3) An additional design, the experimental point at a distant  $\pm \alpha$  from center, while the distance of the axial point was  $\pm 1.68$  (2<sup>n/4</sup> = 1.682 for n = 3) calculated by Equation (2).

$$\alpha = (2^n)^{1/4}$$
 (2),

where  $\alpha$  is the distance of the axial points and *n* is the number of independent variables. The coefficient of the polynomial model was calculated by Equation (3).

$$Y = b_0 + b_1 X_1 + b_2 X_2 + b_3 X_3 + b_{11} X_1^2 + b_{22} X_2^2 + b_{33} X_3^2 + b_{12} X_1 X_2 + b_{23} X_2 X_3 + b_{13} X_1 X_3$$
(3),

where *Y* is the predicted reducing sugar,  $X_1$ ,  $X_2$ ,  $X_3$  are the independent variables,  $b_0$  is the offset term,  $b_1$ ,  $b_2$ ,  $b_3$  are the linear effects,  $b_{11}$ ,  $b_{22}$ ,  $b_{33}$  are the square effects, and  $b_{12}$ ,  $b_{23}$ ,  $b_{13}$  are the cross effects of the interaction terms (Khuri and Mukhopadhyay, 2010; Bezerra *et al.*, 2008).

### 3. Result and Discussion

#### 3.1 Components of corncob

Components of the fresh corncob are shown in Table 1. The major components are 22.76 % carbohydrate that can be hydrolyzed to fermentable sugars before transforming into ethanol and 75.48 % moisture that can support the good growth of microorganisms and save water material used in the fermentation process. This showed that the corncob was a potential material for the ethanol fermentation.

#### 3.2 Response surface analysis for the optimization of three factors

The important factors for this clarification, hydrolysis process, to produce the reducing sugars are time, temperature and alpha-amylase amount. This method evaluates the effects of the hydrolysis process, design model used to study interaction of the three factors and to find the optimum condition. The experimental conditions are shown in Tables 2 and 3. The results for central composite design (CCD) are shown in Table 4, the second-order polynomial equation giving the reducing sugar as a function of time (X<sub>1</sub>,min), temperature (X<sub>2</sub>,°C) and alpha-amylase amount (X<sub>3</sub>,%w) was shown as Equation (4).

$$Y=-100.01+0.061X_{1}+2.235X_{2}+19.90X_{3}-0.0000176X_{1}^{2}-0.01190X_{2}^{2}$$
  
+85.42X\_{3}^{2}-0.000540X\_{1}X\_{2}-0.040X\_{1}X\_{3}-0.339X\_{2}X\_{3} (4),

The RSM predicted and experimental values of the reducing sugar are given in Table 4. To test the fit of the CCD model, the regression equation and determination coefficient ( $R^2$ ) were

estimated. The value of  $R^2$  is 0.906 implied that it was a quite good fit, and that 90.6% of the variation could be explained by the model.

	Iubit		predicted results			
Run no.	$X_1$	X <sub>2</sub>	V	Reducing sugar (g/L)		
Kull IIO.	$\mathbf{MO}. \qquad \mathbf{A}_1 \qquad \mathbf{A}_2$	$X_3$	Experimental	RSM predicted		
1	205	85	0.08	5.64	5.37	
2	150	90	0.13	5.26	5.35	
3	95	95	0.17	5.25	5.31	
4	60	90	0.13	4.82	5.06	
5	150	80	0.13	4.37	4.47	
6	150	90	0.13	5.36	5.35	
7	95	85	0.08	4.76	4.71	
8	150	90	0.20	6.17	6.18	
9	240	90	0.13	5.30	5.35	
10	95	95	0.08	5.18	4.84	
11	95	85	0.17	5.71	5.48	
12	150	100	0.13	3.75	3.85	
13	150	90	0.05	5.20	5.47	
14	205	85	0.17	5.61	5.75	
15	205	95	0.08	4.90	4.93	
16	205	95	0.17	5.15	5.00	
17	150	90	0.13	5.39	5.35	

Table 4: Experimental and RSM predicted results

From Table 5, the fitting model is predicted by the analysis of variance (ANOVA). The ANOVA of the quadratic regression model indicates that the model is highly significant, because of Fisher's F-test (F-model, mean square regression: mean square residual = 7.53) and a very low probability value (P-model > F = 0.00718). As illustrated in Table 6, some effects of factors and their interactions on reducing sugar concentrations are significant (p<0.05) in the ANOVA that indicates a significant effect of the corresponding factors on the response. The p-values from the t-test analysis given in Table 6 are used to determine the significant levels of three process parameters and their interactions on the reducing sugar. The most significant parameter is temperature. The effect of alpha-amylase amount is less significant (p> 0.05) so this interaction can be deleted from Equation (4) without significant effect on the accuracy of predicted reducing sugar concentration. (Yu *et al.*, 2009; Wang *et al.*, 2013).

Tuble et fill (o fill for the full quadrate model						
ANOVA						
Source of variation	%Sum of squares (SS)	Probe > F				
Regression	4.397	91	0.489	7.532	0.00718	
Residual	0.454	9	0.06487			
Total	4.851	100				

 Table 5: ANOVA for the full quadratic model

Term	Coefficient	Value	Standard Error	t -value	P - value
Constant	b0	-100.01	19.81	-5.048	0.00148
Time (min)	b1	0.06053	0.03149	1.922	0.09605
Temperature (°C)	b2	2.235	0.414	5.397	0.00101
Alpha-amylase (%w)	b3	19.90	38.45	0.518	0.621
Time x Time	b4	-1.76 E-05	2.68 E-05	-0.655	0.533
Temperature x Temperature	b5	-0.01190	0.00226	-5.256	0.00118
Alpha-amylase x Alpha-amylase	b6	85.42	38.89	2.196	0.06408
Time x Temperature	b7	-0.000540	0.000334	-1.617	0.150
Time x Alpha-amylase	b8	-0.04026	0.03705	-1.087	0.313
Temperature x Alpha-amylase	b9	-0.339	0.408	-0.830	0.434

Table 6: Coefficients, t-statistics and significant probability of the model for Equation (4).

# 3.3 Interactions among the factors

### 3.3.1 The effects of alpha-amylase amount and temperature

Figure 1 shows the effects of alpha-amylase amount and temperature on reducing sugar content. The reducing sugar content of clarified product increased with increasing amount of alpha-amylase and temperature in the range of 84.4 to 91.1°C. However, the conversion rate was reduced for a further increase in temperature.

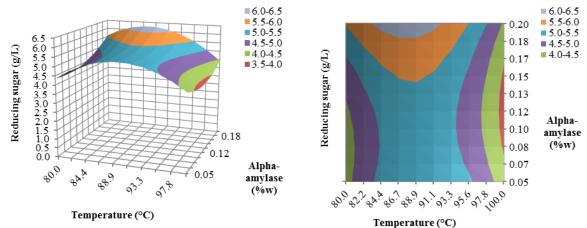


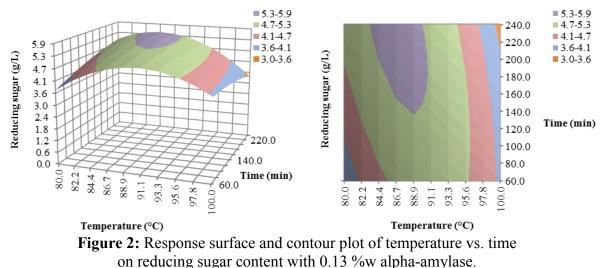
Figure 1: Response surface and contour plot of temperature vs. alpha-amylase on reducing sugar content for 150 min.

## 3.3.2 The effects of heating time and temperature

The effects of heating time and temperature on reducing sugar content are shown in Figure 2. The reducing sugar increased with an increase in both time and temperature. However a higher temperature from 88.9 to 100 °C caused a reduction in the sugar content. To obtain an \*Corresponding author (Sininart Chongkhong). Tel.: +66-7428-7293; fax: +66 7455 8833

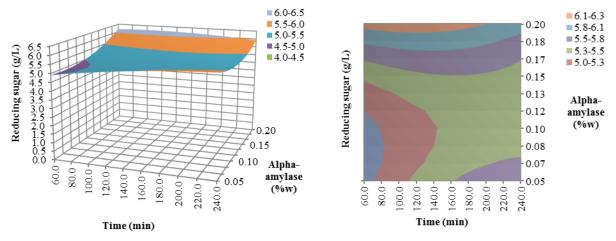


optimum reducing sugar content the clarification process should be operated at a temperature in the range of 84.4 to 88.9°C for a time in the range of 150 to 240 min.



### 3.3.3 The effects of heating time and alpha-amylase amount

The interaction of time and alpha-amylase amount on reducing sugar content (Figure 3) implies that the clarification process should be carried out for a time in the range of 100 to 180 min with 0.17-0.2 %w alpha-amylase to achieve a maximum content of reducing sugar.



**Figure 3:** Response surface and contour plot of time vs. alpha-amylase on reducing sugar content at 90 °C.

The results of the influence and interaction of the factors using CCD indicated that the highest yield could be reached near the center point of the operating conditions as on the contour curves. The optimum condition was at 87.6 °C for 150 min with 0.2 %w alpha-amylase which could provide 6.21 g/L for experimental and 6.25 g/L for predicted reducing sugar contents. These showed that the model, Equation (4), could be useful.

### 4. Conclusion

A clarification step before liquefaction and fermentation steps of the ethanol production from the corncob has been evaluated. The ranges of time, temperature and alpha-amylase amount were established to optimize the operation condition by RSM which could save experimental time and cost. The optimum condition were an alpha-amylase amount of 0.2 %w, a temperature of 87.6 °C and a time of 150 min. that gave the highest amount of 6.21 g/L reducing sugar content.

### 5. Acknowledgement

The authors gratefully acknowledge the financial support from the Graduate school and Faculty of engineering, Prince of Songkla University.

### 6. References

- Agro-Industry department center for export: ADCET. The faculty of Agro-Industry, Prince of Songkla University HatyaiSongkhla 90112.
- Aiyer, P.V. (1995). Amylases and their applications. *African Journal of Biotechnology*, 4(13), 1525-1529.
- Balata, M., Balat, H. and Oz, C. (2008) Progress in bioethanol processing. *Progress in Energy and Combustion Science*, 34(5), 551-573.
- Bandaru, V.V.R., Somalanka, S.R., Mendu, DR., Madicherla, N.R. and Chityala, A. (2006). Optimization of fermentation conditions for the production of ethanol from sago starch by co-immobilized amyloglucosidase and cells of Zymomonasmobilis using response surface methodology. *Enzyme and Microbial Technology*, 38(1-2), 209-214.
- Bezerra, M.A., Santelli, R.E., Oliveira, E.P., Villar, L.S. and Escaleira, L.A. (2008). Response surface methodology (RSM) as a tool for optimization in analytical chemistry. *Talanta*. 76(5), 965-977.
- Chena, M., Xia, L. and Xue, P. (2007). Enzymatic hydrolysis of corncob and ethanol production from cellulosic hydrolysate. *International Biodeterioration& Biodegradation*, 59(2), 85–89.
- Chongkhong, S., Lolharat, B. and Chetpattananondh, P. (2012). Optimization of Ethanol Production from Fresh Jackfruit Seeds Using Response Surface Methodology. *Journal* of Sustainable Energy & Environment, 3, 97-101.
- Das, S., Singh, S., Sharma, V. and Lalsoni, M. (2011). Biotechnological applications of industrially important amylase enzyme. *International Journal of Pharma and Bio Sciences*, 2(1), 486-496.

Khuri, A. and Mukhopadhyay, S. (2010). Response surface methodology. Wiley

<sup>\*</sup>Corresponding author (Sininart Chongkhong). Tel.: +66-7428-7293; fax: +66 7455 8833 E-mail addresses: <u>csininart@yahoo.com</u>. © 2014. International Transaction Journal of Engineering, Management, & Applied Sciences & Technologies. Volume 5 No.1 ISSN 2228-9860 eISSN 1906-9642. Online available at <u>http://tuengr.com/V05/0067.pdf</u>

Interdisciplinary Reviews-Computational Statistics, 2, 128-149.

- Kreuger, E., Sipos, B., Zacchi, G., Svensson, S. and Björnsson, L. (2011). Bioconversion of industrial hemp to ethanol and methane: The benefits of steam pretreatment and co-production. *Bioresource Technology*, 102(3), 3457-3465.
- Liu, K., Lin, X., Yue, J., Li, X., Fang, X., Zhu, M., Lin, J., Qu, Y. and Xiao, L. (2010). High concentration ethanol production from corncob residues by fed-batch strategy. *Bioresource Technology*, 101(13), 4952-4958.
- Lolharat, B., Chongkhong, S. and Chetpattananondh, P. (2011). Optimizing conditions for enzymatic clarification of prebiotic extracted jackfruit seeds using response surface methodology. *Proceeding of the 5<sup>th</sup> International Conference on Engineering and Technology (ICET-2011)*, May 2-3, 2011, Phuket, Thailand.
- Miller, G.L. (1959). Use of dinitrosalicyclic acid reagent for determination of reducing sugar. *Analytical Chemistry*, 31(3), 426-428.
- Reed, G. and Nagodawithana, T.W.(1995). Enzyme, Biomass, Food and feed. *Biotechnology*, 9(2),676.
- Wang, Q., Ma, H., Xu, W., Gong, L., Zhang, W. and Zou, D.(2008). Ethanol production from kitchen garbage using response surface methodology. *Biochemical Engineering Journal*, 39(3), 604-610.
- Wang, L., Luo, Z. and Shahbazi,(2013). A. Optimization of simultaneous saccharification and fermentation for the production of ethanol from sweet sorghum (Sorghum bicolor) bagasse using response surface methodology. *Industrial Crops and Products*, 42, 280-291.
- Yu, J., Zhang, Xu. and Tan, T. (2009). Optimization of media conditions for the production of ethanol from sweet sorghum juice by immobilized *Saccharomyces cerevisiae*. *Biomass* and *Bioenergy*, 33(3), 521-526.
- Zhu, S., Wu, Y., Yu, Z., Chen, Q., Wu, G., Yu, F., Wang, C. and Jin, S. (2006). Microwave-assisted Alkali Pre-treatment of Wheat Straw and its Enzymatic Hydrolysis. *Biosystems Engineering*, 94(3), 437-442.



**Dr.Sininart Chongkhong** is an Assistant Professor of Department of Chemical Engineering at Prince of Songkla University. She received her B.Eng. from Prince of Songkla University with Honors in 2002. She continued her Ph.D. study at Prince of Songkla University, where she obtained her Ph.D. in Chemical Engineering. Dr. Sininart Chongkhong currently works on ethanol/biodiesel technologies.



**Woraluk Kongjindamunee** holds a degree in Chemical Engineering from Prince of Songkla University, Thailand. She is interested in applications of a green chemical technology.

**Peer Review:** This article has been submitted, peer-reviewed, and awarded best paper from the Third International-Thai Chemical Engineering and Applied Chemistry (TIChE) Conference, jointly organized by Department of Chemical Engineering, Faculty of Engineering, Khon Kaen University and Thai Institute of Chemical Engineering and Applied Chemistry, at Pullman Khon Kaen Raja Orchid Hotel, Khon Kaen, THAILAND, October 17-18, 2013.