



## Effect of Laser Priming on accumulation of Free Proline in Spring Durum Wheat (*Triticum turgidum* L.) under Salinity Stress

Naemeh Zare <sup>a</sup>, Seyed Ahmad Sadat Noori <sup>a\*</sup>, Nayer Azam Khosh kholgh Sima <sup>b</sup>, and Seyed Mohammad Mahdi Mortazavian <sup>a</sup>

<sup>a</sup> Department of Agronomy and Plant Breeding, College of Abouraihan, University of Tehran, Tehran, Iran

<sup>b</sup> Agricultural Biotechnology Research Institute of Iran, Karaj, Iran

### ARTICLE INFO

#### Article history:

Received 01 November 2013

Received in revised form 12 December 2013

Accepted 16 December 2013

Available online 20 December 2013

#### Keywords:

*laser irradiation;*  
*proline accumulation;*  
*salt sensitive;*  
*salt tolerance;*  
*seed treatment technique.*

### ABSTRACT

Experiments were conducted during 2012 in a greenhouse of College of Abouraihan, University of Tehran, Iran. In this study the effects of salt stress and laser priming on proline content of durum wheat (*Triticum turgidum* L.) was carried out in a factorial experimental based on a randomized complete block design (RCBD) with three replications. Seeds from two cultivars salt sensitive and salt tolerance of Durum wheat (*Triticum turgidum* L.) were exposed to neodymium-doped yttrium aluminum garnet (Nd-Yag) laser irradiation (75 mW cm<sup>-2</sup>, radiated for 12 min). Salinity treatments carried out in four levels (Control, 70, 140 and 210 mM) via sodium chloride. The sampling from first leaves was carried out on four stages of growth and their proline content was measured. The result showed that free proline content in leaves increased significantly by increasing of NaCl concentration and salt tolerant variety accumulate more Proline than sensitive variety. Also proline content significantly increased with irradiation by laser beam in salinity condition. These results indicate that the low power continuous wave Nd-Yag laser light seed treatment has considerable biological effects on plant metabolism. This seed treatment technique can be potentially employed to enhance agricultural productivity.

© 2014 INT TRANS J ENG MANAG SCI TECH.



## 1. Introduction

Environmental stresses negatively influence the plant growth, developmental stage and crops yield. Soil salinity is one of the major abiotic stresses adversely affects physiological and metabolic processes such as germination percentage, crop growth, productivity and photosynthesis in plants (Sairam *et al.*, 2002). Soil salinity makes the change in plants in two ways. High concentration of salts in the soil makes it harder for roots to absorb water and leads to physiological drought, and in plant can be toxic by high concentration of  $\text{Na}^+$ . Another way is that the salts on the outside of roots have an immediate effect on cell growth and associated metabolism; toxic concentrations of salts take time to accumulate inside plants before they affect plant function (Khosh KholghSima *et al.*, 2009; 2012). Because NaCl is the most soluble and widespread salt, it is not surprising that all plants have evolved mechanisms to regulate its accumulation and to select against it in favor of other nutrients commonly present in low concentrations, such as  $\text{K}^+$  and  $\text{NO}_3^-$  (Munns, and Tester, 2008). If  $\text{Na}^+$  and  $\text{Cl}^-$  are sequestered in the vacuole of a cell, organic solutes that are compatible with metabolic activity even at high concentrations (hence ‘compatible solutes’) must accumulate in the cytosol and organelles to balance the osmotic pressure of the ions in the vacuole (Flower *et al.*, 1977. Wyn Jones *et al.*, 1977). The compounds that accumulate most commonly are sucrose, proline, and glycine betaine, although other molecules can accumulate to high concentrations in certain species (Hasegawa Pm *et al.*, 2000. Munns, 2005), Production and accumulation of Free Amino Acids (FAA), especially proline by plant tissue during drought, salt and water stress is an adaptive response. This amino acid is widely believed to function as a protector or stabilizer of enzymes or membrane structure that are sensitive to dehydration or ionically induced damage such as Reactive Oxygen Species (ROS) and antioxidant defense. For the durum subspecies, high the level of free amino acids, especially proline in the leaf correlated well with salinity tolerance (simon-sarkadi et al, 2002). Durum wheat (*Triticum durum* Desf), which is used mainly for making pasta and macaroni is the second most important wheat, and is widely grown in Southern Europe and the Middle East, and on soils affected by salinity (Sadat Noori and McNeilly, 2000). Durum wheat compared with common bread wheat (*Triticum aestivum* L.) is known for its hardness, protein, intense yellow color, nutty flavor and excellent cooking qualities (Kneipp, 2008). Although durum wheat cultivars are more salt sensitive than bread wheat and their yield is lower under saline soils (Munns and James, 2003), for this reason breeding new cultivars of durum wheat capable of that can be grown on

saline soils is of great interest. Seed dressing with various growth regulators, plant hormones, fertilizers etc. are currently considered the most efficient; the best recognized and the most often used practice. However, such substances may modify the chemical structure of the treated seeds, pollute the soil and pose a great danger to the environment. Therefore, more attention has been paid to study physical factors that favorably improve cultivated plants (Perveen, R., et al. 2010). Many studies indicate that physical methods stimulate only changes at physiological and biochemical level in the treated seeds (Aladjadjiyan, 2007; Perveen *et al.* 2010) rendering them safe and friendly to the environment. Laser is considered one of the physical methods that can be safely applied to improve the quality and yield of crop plants (Inyushin, *et al.*, 1981; Ivanova, 1998; Koper, 1994; Podleony, 2002). The aim of these methods is the appropriate preparation of the sowing material to improve seed sprouting growth and vigor (Podlesny, and Podlesna, 2004) The laser radiation has been used by different researches (Wilczek, *et al.* 2004; Chirkova, 2002; Podlesny, Podlesna, 2004) as a physical method to improve the germination, the growth and the vigor of seeds (Salyaev *et al.* 2007). Biophysical methods can stimulate the seed and plants, through improving the energy balance and hence activating the growth and yield processes (Chen *et al.*, 2005; Vasilevski, 2003). However, biophysical protocols are beneficial that enable plants to vegetate at a higher energy level. It is now evident that physical methods such as laser radiation application enhance the energy account of metabolites by internal energy transformation (Chen, *et al.* 2005). Therefore; the aim of the present study was to investigate the effect of pre sowing laser treatment on the accumulations of free proline content under salinity stress.

## 2. Mathematical Model

Experiments were conducted during 2011 in a greenhouse of College of Abouraihan, University of Tehran, was carried out using a factorial (salinity, laser, cultivar) based on a completely randomized block design with three replications. Factors included four salinity levels (Control, 70, 140, 210 mM NaCl), two cultivars (*Triticum turgidum* L. cv. Karkheh and Dena) and two lasers level (irradiated seeds and Non-irradiated). Seeds from two cultivars salt sensitive (Karkheh) and salt tolerance (Dena) of Durum wheat (*Triticum turgidum var.durum* L.) were obtained from Seed and Plant Improvement Institute in Karaj, Iran. Primary the selection of seeds based on their sizes was carried out and irradiated with Neodymium-Yttrium-Aluminum Garnet (Nd:YAG) laser (wavelength 532nm, power intensity 75 mW cm<sup>-2</sup>

and irradiation time for 12 minutes). The irradiated seeds sown in a 48 plastic pots that have Seven kilogram of soil composed of dried sandy loam with natural pH (pH=7.28). The pots were placed in a green house in semi-controlled conditions with a photoperiod of 16h light and 8h dark, relative humidity of 60%-75%. When seedling were in zadoks 21, salt stress treatments were imposed by adding 70, 140 and 210mM NaCl by adjustment the water content of soil to near the field capacity. The sampling from first leaves was carried out after 24 half, 3 days, 7 days and 15 days of each salinity level, then incubated in liquid nitrogen and maintained at -80°C until the measurement time.

The proline content was quantified according to Bates *et al.* (1973). Leaf samples (0.2–0.5 g of fresh weight) of frozen plant material were ground to a fine powder in a pre-cooled mortar with liquid nitrogen. The powder was homogenized in 5 mL of 3% aqueous sulfo-salicylic acid and centrifuged at 14000g for 2 minutes. Two mL of acid-ninhydrin and 2 mL of glacial acetic acid were mixed with 2 mL of the homogenate in a test tube. The mixture was incubated at 100°C for an hour. Reaction was stopped by placing the test tube in an ice bath. Four mL of toluene were added to each test tube and vortexed for 15–20 seconds. The organic and inorganic phases were separated, and the absorbance at 520 nm of the organic toluene phase containing the chromophore was recorded with spectrophotometer (Perkin-Elmer, Lambda 25, USA). Concentrations of proline in plant tissue are expressed on a fresh weight basis and determined from a standard curve and calculated on a fresh weight basis as follows:

$$\mu\text{moles} \frac{\text{Proline}}{\text{g}} \text{ of fresh weight material} = \frac{\left[ \frac{\mu\text{g-Proline} \times \text{ml Toluene}}{\text{ml}} \right]}{115.5 \frac{\mu\text{g}}{\mu\text{mole}}} \quad (1).$$

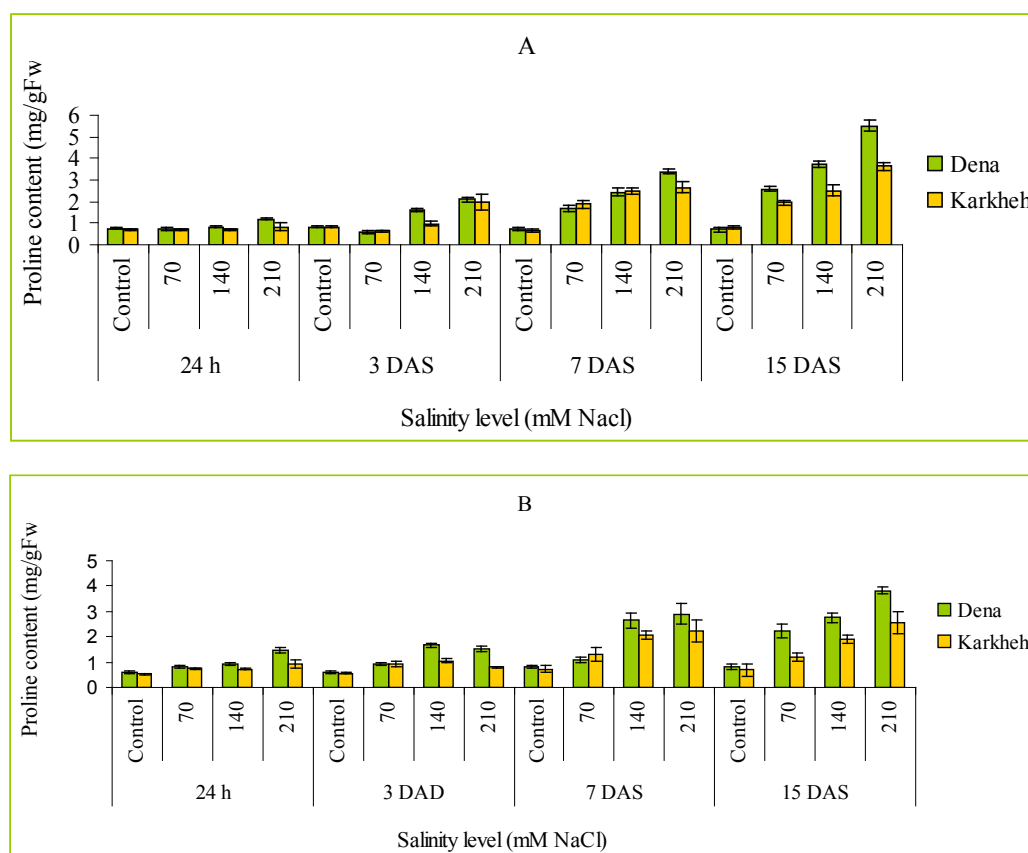
### 3. Statistical Analyses

Data were subjected to analysis of variance (ANOVA) using the general linear model of SAS (Statistical Analysis System V.9) program. The mean differences were compared by Duncan's test at the  $P \leq 0.05$  and 0.01 levels.

### 4. Results and Discussion

The results of analysis variance of effects of laser pretreatment and salinity stress on proline accumulation were showed in Table 1. As we can see, the interaction effect of salinity

and laser on free proline content was significant in all steps on sampling ;containing 24h, 3days, 7days and 15 days after salinization (DAS).



**Figure 1:** Effect of salinity levels on proline in durum wheat genotypes with Effect of Laser (A) and No Effect of Laser (B). Vertical bars indicate  $\pm$  S.E. of mean (n=3). Data were significant at 5%probability level for days, salinity treatments and varieties.

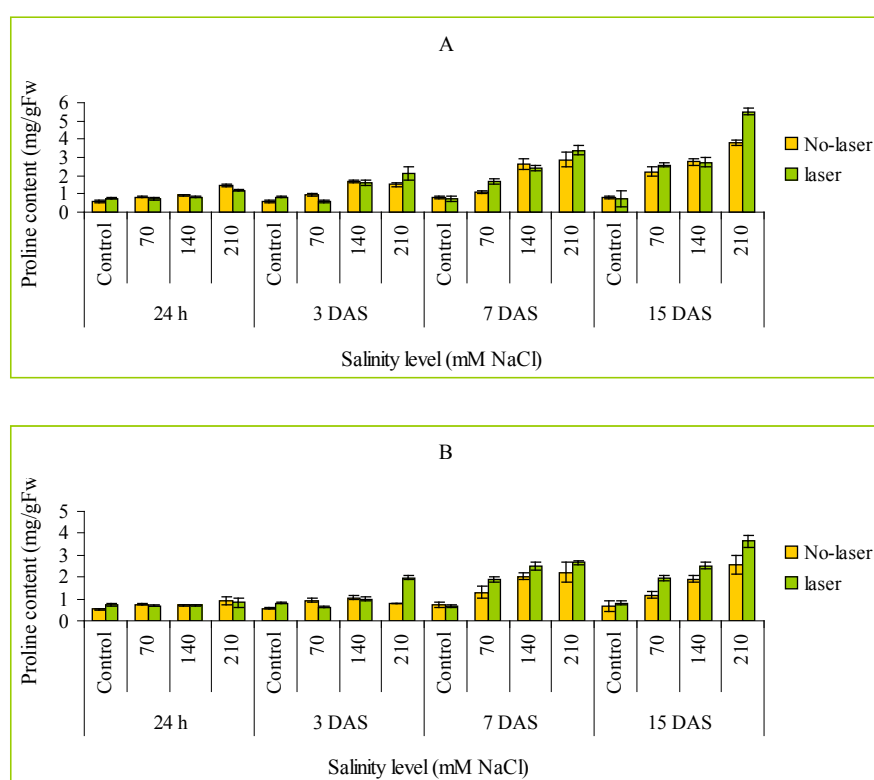
**Table 1:** Analysis of variance for proline content of durum wheat (*Triticum turgidum* L.) was irradiated with laser under salinity conditions

Source of variation	df	Mean of squares			
		24h	3d	7d	15d
Replication	2	0.087**	0.192**	1.164**	0.76**
Variety	1	0.373**	0.850**	0.548**	8.97**
Salinity	3	0.440**	2.253**	10.209**	20.66**
Laser	1	0.012 <sup>n.s</sup>	0.394**	0.916**	5.69**
Variety $\times$ Salinity	3	0.107**	0.292**	0.426**	1.25**
Variety $\times$ Laser	1	0.023 <sup>n.s</sup>	0.073 <sup>n.s</sup>	0.068 <sup>n.s</sup>	0.02 <sup>n.s</sup>
Salinity $\times$ Laser	3	0.065*	0.807**	0.274**	0.95**
Variety $\times$ Salinity $\times$ Laser	3	0.003 <sup>n.s</sup>	0.066 <sup>n.s</sup>	0.089 <sup>n.s</sup>	0.1 <sup>n.s</sup>
Error	30	0.016 <sup>n.s</sup>	0.029 <sup>n.s</sup>	0.07 <sup>n.s</sup>	0.07 <sup>n.s</sup>
C. V. (%)	--	16.53	18.59	19.69	15.18

\*, \*\*significant at 5% and 1%, respectively. ns = non-significant.

Salinity caused an important increase in proline content in both genotypes. Proline content also increased very significantly at the 3th stage as compared to the 1st and 2nd stage and then marginally at the 4th stage in both cultivars. In both conditions, laser and No-laser, two genotypes had not significant differences at low solution of salinity (70 and 140 mM) in 24h after doing of salinity stress. This revealed that differences between studied genotypes based on proline content, are obvious only after some time and in higher solution of salinity. In present of laser and at 210 mM, 'Dena' showed more proline content than 'Karkheh' statistically at all stages except of 3days after doing of salinity stress (Figure 1.A). Maximum differences between two genotypes were observed at 210 mM NaCl concentrations at the last stage of sampling. Proline content in plant cells under salt stress is a universal phenomenon that can serve as an osmotic regulator (El-Sayed *et al.* 2007) and is widely documented in the cell pressure adjustment, detoxification of injurious ions and membrane stabilization in plants under salinity conditions (Ashraf and Foolad 2007). Proline has been shown to function as a molecular chaperone able to protect protein integrity and enhance the activities of different enzymes (La' szlo' Szabados and ArnouldSavoure'. 2009). In No-laser condition, differences between genotypes were more obvious at starting stages of sampling, 3days, in contrast to laser condition. In study of two genotypes that were not exposed to laser, it was resulted that proline content of 'Dena' was more than 'Karkheh' statistically in all levels of salinity at 7day and 15 day except 70 level of salinity at 7day (Figure 1.B). In the 3rd stage of sampling, differences between genotypes showed that at 140 and 210 mM NaCl 'Dena' had 22% and 23% more proline content, respectively, comprising to the control. In the 4th stage and 140 mM NaCl so 'Dena' had 30% more pro than 'Karkheh' and at the 210 mM concentration 32%. During our investigation, analysis of variance showed that with the increase of salinity irrigation, proline content increased; namely 210 mM NaCl induced the highest value and control sample had the minimum extent of proline content in two varieties (Figure 1). Effect of salinity to proline content in canola, rice and wheat was reported previously (Shamseddin-Saeid and Farahbakhsh, 2008; Azizpour *et al.* 2010; Hadi *et al.* 2007). Expression of the genes encoding cell wall proteins (proline-rich protein and extension) and cellulose synthesis was induced in barley roots by salt stress (Ueda *et al.* 2007). Enzymes of the ROS-scavenging glutathione-ascorbate cycle showed significantly lower activities in the p5cs1 mutants compared to wild type under salt stress suggesting that Pro accumulation is implicated in the control of either stability or activity of enzymes in the glutathione-ascorbate cycle (Sze'kely *et al.* 2008). There are some reports showing that pretreatment of seeds by

laser beams increased the quality and quantity of produced plants. According to Cwintalet al. (2010), presuming stimulation of seeds with laser light caused a significant increase in the content of specific proteins, phosphorus and molybdenum in dry matter of the plants, and a decrease in the content of crude fiber. In our research, laser priming caused a reduction of undesirable effects of salinity and an increase in the proline content of plants under both normal and stress conditions. With the comparison of proline content between control and plants irradiated with laser beam in the most of same salinity concentration, we observed that laser had a significant effect ( $P < 0.001$ ) on proline content under salinity condition (Figure 2).



**Figure 2:** Effect of Laser Irradiation on proline content in durum wheat genotypes; salt tolerant (A) and salt sensitive(B). Vertical bars indicate $\pm$ S.E. of mean (n=3). Data were significant at 5% probability level for days, salinity treatments and varieties.

As we can see on Figure 2A, there was not any significant difference between salinity levels at 24h stage in all salinity levels except of 70 mM. In Karkheh (a sensitive genotype), differences among using of laser and No-laser are completely visual in higher dose of salinity, especially after 7 days stage. In Dena (a tolerance genotype) and in 210 mM NaCl, using of



laser increased the proline content statistically after 3 days, 7 days and 15 days of salinity stress, while it was reduced in 24 days stage comprising to No-laser treatment (Figure 2.B). Also, using of laser in 140 mM NaCl of salinity was not change significantly the Dena's proline content in all stage except of 24h stage that it was reduced significantly in laser condition.

Proline was shown to protect Complex II of the mitochondrial electron transport chain during salt stress and therefore stabilized mitochondrial respiration (Hamilton et al, 2001). There are some reports that shows laser pretreatment had a positive effect on proline accumulation in canola (Ashrafijou et al 2010) and common wheat (Sadat Noori et al, 2011) lead to increase salinity tolerance. One of the reasons for proline content increase can be the additional energy in plant at irradiation with laser beams. The laser Beam, as specific light, can be absorbed effectively through the macromolecules and cause some photochemical impacts (Xiang, 1995). In our research, the highest content of proline was observed in high salinity's stress, 210 mM NaCl, in Dena genotype that irradiated with laser, so proline content can a good choice for indirect selection in breeding programs for tolerance to salinity stress and also, the laser use can an alternative way for this purpose. It seems that the effects of salinity on plant growth and its osmolytes are not high in start stage of growth and sampling for examination of its effects on plants can be carry out on late period of plant's life cycle.

## 5. Conclusion

1. Increasing of the salinity, increases proline content and 210 mM NaCl induced the highest amount of this amino acid.
2. Salt Tolerant variety accumulates free proline content more than salt sensitive variety, so proline accumulation can be a useful parameter to breeding program for salinity tolerance.
3. Laser irradiation increased proline content in most of salinity levels and sampling steps.

The result showed that free proline content in leaves increased significantly by increasing of NaCl concentration and salt tolerant variety accumulate more Proline than sensitive variety. Also proline content significantly increased with irradiation by laser beam in salinity condition. These results indicate that the low power continuous wave Nd-Yag laser light seed treatment has considerable biological effects on plant metabolism. This seed treatment technique can be potentially employed to enhance agricultural productivity. From this study,



stimulating effect of laser radiation can be used in wheat breeding and investigating the use of laser beam on other plants is recommended.

## 6. Acknowledgements

The authors are highly thankful to Dr. Mohsen Esmaeel Zadeh Moghadam, Wheat Breeder of Seed and Plant Improvement Institute, for supplying the seeds of durum wheat genotypes and Dr. Mohammad Ali Ansari from Laser and Plasma Research Institute of Shahid Beheshti University.

## 7. References

- Aladjadjiyan A. 2007. The use of physical methods for plant growing stimulation in Bulgaria. *Journal of Central Eurpian Agriculture*, 8, 3: 369-380
- Amirjani M.R. 2010. Effect of Salinity Stress on Growth, Mineral Composition, Proline Content, Antioxidant Enzymes of Soybean. *American Journal of Plant Physiology*, 5, 6: 350-360
- Ashrafijou M., Sadat Noori S.A., Izadi Darbandi A., Saghafi S. 2010. Effect of salinity and radiation on proline accumulation in seeds of canola (*Brassica napus* L.), plant soil environment, 56, 7: 312–317
- Ashraf M., Foolad M.R. 2007. Roles of glycine betaine and proline in improving plant abiotic stress resistance. *Environmental and Experimental Botany*, 59: 206–216
- Azizpour. K., Shakiba M.R., Khosh Kholgh Sima N.A., Alyari H., Moghaddam M., Esfandiari E. and Pessarakli. M. 2010. Physiological response of spring durum wheat genotypes to salinity. *Journal of Plant Nutrition*, 33:859–873
- Chen Y.P., Yuea M. and Wang X. L. 2005. Influence of He–Nelaser irradiation on seeds thermodynamic parameters and seedlings growth of *Isatis indogotica*. *Plant Science*, 168: 601–606
- Chen Y.P., liu Y.J., Wang X.L., Ren Z.Y. and Yue M. 2005. Effect of microwave and He-Ne laser on enzyme activity and biophoton emission of *Isatisindigotica* Fort. *Journal of Integrative Plant Biology*, 47, 7: 849–855
- ChirkovaT.V. 2002. Physiological mechanism of stability in plants. St. Petersburg, S.-Peterb. U.
- C' wintal M., Dziwulska-Hunek A. and Wilczek M. 2010. Laser stimulation effect of seeds on quality of alfalfa. *International Agrophysics*, 24:15–19
- FAO. 2008. FAO Land and Plant Nutrition Management Service.

<http://www.fao.org/ag/agl/agll/spush>.

- Flowers T.J. and Yeo A.R. 1995. Breeding for salinity resistance in crop plants. *Australian Journal of Plant Physiology*, 22:875–884.
- Flowers T.J., Troke P.F. and Yeo A.R. 1977. The mechanism of salt tolerance in halophytes. *Annual Review of Plant Physiology*. 28:89–121.
- Hadi M.R., Khavarinejad R. and Khosh kholgh sima N.A. 2007. NaCl Effect on accumulation of Minerals ( $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Cl}^+$ ) and proline in *tritucum turjidum* L. *Asian journal of plant science* 6,2: 379-383.
- Hamilton E.W. and Heckathorn S.A. 2001. Mitochondrial adaptations to NaCl complex I is protected by anti-oxidants and small heat shock proteins, whereas complex II is protected by proline and betaine. *Plant Physiology*, 126: 1266–1274.
- Hasegawa P.M., Bressan R.A., Zhu J.K. and Bohnert HJ. 2000. Plant cellular and molecular responses to high salinity. *Annual Review of Plant Physiology and Plant Molecular Biology*.51: 463–99.
- Hernández A., Carballo C.A., Artola A. and Michtchenko A. 2006. Laser irradiation effects on maize seed file performance, *Journal Seed Science and Technology*, 34, 1: 193-197
- Inyushin W.M., Iliasor G.U. and Fedorowa N.N. 1981. *Laser Light and Crop*. Kainar Publication, Ama-Ata.
- Ivanova, R. 1998. Influence of pre-sowing laser irradiation of seeds of introduced flax varieties of linseed oil on yield quality. *Bulgarian Journal of Agriculture Science*, 4: 49-53.
- Kneipp J. 2008. Technical Specialist, Farming Systems. NSW Department of Primary Industries, Tamworth Agricultural Institute, Calala State of New South Wales through NSW Department of Primary Industries.
- Koper R. 1994. Pre-sowing laser biostimulation of seeds of cultivated plants and its results in agrotechnics. *International Agrophysics*, 8, 593-596.
- Khosh Kholgh Sima N. A., Tale A. S., Alitabar R. A, Mottaghi A., Pessarakli, M. 2012. Interactive effects of salinity and phosphorus nutrition on physiological responses of two barley species. *Journal of Plant Nutrition*, 35:1411–1428.
- Khosh Kholgh Sima, N. A., Askari, H., Mirzaei, H. H., and Pessarakli, M. 2009. Genotype-dependent differential responses of three forage species to Ca supplement in saline conditions. *Journal of Plant Nutrition*, 32: 579–597.
- La' szlo' Szabados and ArnouldSavoure'. 2009. Proline: a multifunctional amino acid *Plant Science* Vol.15 No.2. <http://dx.doi.org/10.1016/j.tplants.11.009>.
- Bates L.S., Waldren R.P. and Teare I.D. 1973. Rapid determination of the free proline in water stress studies. *Plant Soil*, 39: 205–208.
- Rensburg L., Krüger G.H.J. and KrügerH. 1993. Proline Accumulation as Drought-tolerance

- Selection Criterion: its Relationship to Membrane Integrity and Chloroplast Ultra structure in *Nicotian atabacum* L. *Journal Plant Physiology*, 141: 188-194.
- Munns R. 2005. Genes and salt tolerance: bringing them together. *New Phytology*. 167:645–63
- Munns R. and James RA. 2003. Screening methods for salinity tolerance: a case study with tetraploid wheat. *Plant Soil*, 253: 201–218.
- Munns., R. and Tester., M. 2008. Mechanisms of Salinity Tolerance, *Annual Review of Plant Biology*, 59: 651–81.
- Perveen R., Ali Q., Ashraf M., Al-Qurainy F., Jamil Y. and Raza A.M. 2010. Effects of Different Doses of Low Power Continuous Wave He–Ne Laser Radiation on Some Seed Thermodynamic and Germination Parameters, and Potential Enzymes Involved in Seed Germination of Sunflower (*Helianthus annuus*L.). *Photochemistry and Photobiology*, 86: 1050–1055
- Podlesny J., Podlesna A. 2004. Morphological changes and yield of selected species of leguminous plants under the influence of seed treatment with laser light. *International Agrophysics*. 18, 3: 253-260
- Podleoeny J. 2002. Studies on influence of laser light on seeds, growth, development and yielding of the white lupine (*Lupinus albus*L.) plants (in Polish). *Monografie I Rozprawy Naukowe, IUNG, Puławy*, 3: 1-59
- Sadat Noori S.A. and McNeilly T. 2000. Assessment of variability in salt tolerance based on seedling growth in *Triticum durum* Desf. *Genetic Resources and Crop Evolution* 47: 285–291
- Sadat Noori, S. A., Ferdosizadeh, L., Izadi Darbandi., A., Mortazavian., S. M. M., Saghafi., S. 2011. Effects of Salinity and Laser Radiation on Proline Accumulation in Seeds of Spring Wheat. *Journal of Plant Physiology and Breeding*. 1(2): 11-20.
- Sairam R.K., Veerabhadra Rao K. and Srivastava G.C. 2002. Differential response of wheat genotypes to long term salinity stress in relation to oxidative stress, antioxidant activity and osmolyte concentration. *Plant Science*, 163: 1037-1046.
- Salyaev R.K., Dudareva L.V., Lankevich S.V., Makarenko S.P., Sumtsova V.M. and Rudikovskaya E.G. 2007. The effect of Low-intensity laser irradiation on the chemical composition and structure of lipids in wheat tissue culture. *Dokl. Akad. Nauk.*, 412: 87-88.
- Simon-sarkadi L.S., Kocsy G. and Sebestyén Z. 2002. Effect of salt stress on free amino acid and polyamine content in cereals. *Proceeding of the 7<sup>th</sup> Hungarian Congress on plant Physiology*, 46: 73-75.
- Shamseddin-Saeid M., Farahbakhsh H. 2008. Investigation of quantitative and qualitative parameters of canola under salty conditions for determining the best tolerance index. *JWSS - Isfahan University of Technology*. 12, 43: 65-78.

Sze'kely G., Abraham E., Cseplo A., Rigo G., Zsigmond L., Csiszar J., Ayaydin F., Strizhov N., Jasik J., Schmelzer E., Koncz C. and Szabados L. 2008. Duplicated P5CS genes of Arabidopsis play distinct roles in stress regulation and developmental control of proline biosynthesis. *Plant Journal*, 53: 11–28.

Ueda A., Yamamoto-Yamane Y. and Takabe T. 2007. Salt stress enhances proline utilization in the apical region of barley roots. *Biochemical and Biophysical Research Communications*: 355, 61–66.

Vasilevski, G. 2003. Perspectives of the application of biophysical methods in sustainable agriculture. *Bulg. Journal of Plant Physiology*, special issue: 179–186.

Wilczek M., Koper R., Cwintal M. and Kornilowicz-Kowalska T. 2004. Germination capacity and the health status of red clover seeds following laser treatment. *International Agrophysics*, 18, 3: 289-293.

Wyn Jones R.G., Storey R., Leigh R.A., Ahmad N. and Pollard A. 1977. A hypothesis on cytoplasmic osmoregulation. In *Regulation of Cell Membrane Activities in Plants*, ed. E Marr'e, O Cifferi, Amsterdam, Netherlands: Elsevier: 121–136.

Xiang, Y. 1995. *Laser Biology*, Hunan Science and Technology Press, Changsha: 124-127.



**Naemeh Zare** obtained her MSc from University of Tehran in 2012. Her thesis is about application Laser Radiation as seed priming technique to increase plant tolerance under stress condition especially salinity, and evaluation this treatment by measuring germination factors, physiological traits and yield and its components in durum wheat under salinity condition.



**Dr. Seyed Ahmad Sadat Noori** earned his Ph.D. in Genetics and Plant Breeding from the University of Liverpool, UK in 1999. He is lecturer of subjects such as Genetics, Advance genetics, Molecular genetics, Plant breeding, Statistics, Population genetics, Agricultural experimental design, in University of Tehran. He published 7 books on these subjects. He is currently full Professor at University of Tehran. His work focuses on salinity tolerance in crop plants (wheat, rice, canola) and medical plants (*Carum copticum* L.) using traditional and novel techniques.



**Dr. Nayer Azam Khoshkholgh Sima** received her PhD in Plant Molecular Physiology from University of Hiroshima, Japan. She is currently working as a Research Assist. Professor at Agricultural Biotechnology Research Institute of Iran (ABRII) and board member of Crop Science and Biotechnology Associations of Iran. Dr. Khoshkholgh Sima's broad research focus is plant stress physiology and phyto/bio remediation. She is currently serving as the editor for the *Journal of Plant Molecular Breeding*, *Iranian Journal of Genetics and Plant* and *Journal of Biosafety*. Dr. Khoshkholgh Sima has been working closely with Ministry of Agriculture of Iran in her capacity as the co-founder of ABRII and with Iran Biotechnology Information Center (IRBIC) for the promotion of biotechnology in Iran.



**Dr. Seyed Mohammad Mahdi Mortazavian** is Assistant Professor at the University of Tehran, college of Aburaihan, Pakdasht. In 2004, he received his MS in plant breeding and in 2009 his PhD in plant molecular breeding and genetic engineering at University of Tehran. Dr. Mortazavian passed a sabbatical leave in WUR, Netherlands for a year. He began as an Editor with *The Journal of applied breeding of crop and horticultural plants* in 2012. His research focuses on cereal (wheat and barley) and medicinal plant (cumin) breeding using traditional and novel techniques.

**Peer Review:** This article has been internationally peer-reviewed and accepted for publication according to the guidelines in the journal's website.